

UNIVERSITAT AUTÒNOMA DE BARCELONA

**Facultat de Medicina i Cirurgia
Departament de Medicina**



**APLICABILITAT CLÍNICA DE LES TÈCNIQUES DE
DETECCIÓ *IN VITRO* DE L'INTERFERÓ-GAMMA EN LA
INFECCIÓ I LA MALALTIA TUBERCULOSA**

Maria Luiza de Souza Galvão

2014

cannot be extrapolated to countries with moderate or low TB prevalence. Indeed, a decreased T-cell response against ESAT-6 stimulation was reported in household contact children exposed to *Mycobacterium africanum* (responsible for up to half of TB cases in western Africa), rather than that observed in children exposed to *M. tuberculosis* [89].

Regarding the specificity of tests in NTM infection, Detjen *et al.* analyzed 23 children with bacteriological NTM lymphadenitis and 22 with other nonmycobacterial respiratory tract infections, and reported a specificity for the T-SPOT.TB and QFN-G-IT tests of 100% [90]. Although the specificity of IFN- γ -based tests is excellent, frequent discordant results with the TST have been described [72,73,76,77,83,84,91–93]. Among 511 TST-positive children screened for LTBI at school (14–15 years old) in Norway [84], only 44 (9%) had a confirmed positive QFN-G result. In another study conducted in Barcelona (Spain) among children who were not BCG-vaccinated, QFN-G-IT was negative in 60.4% of children with a positive TST, and T-SPOT.TB assay in 56.6% of cases [76]. In fact, a priority research area is to understand discordant TST and IFN- γ -based test results, including the role of NTMs [11].

With regard to indeterminate results, Ferrara *et al.* noted that QFN-G obtained in children under 5 years gave a higher proportion of indeterminate results than the T-SPOT.TB (32 vs 0%) [12]. A higher ratio of QFN-G-indeterminate results (17%) was also obtained by Connell *et al.* [72]. However, in a later study, Connell *et al.* reported a higher proportion of indeterminate results by the T-SPOT.TB than the QFN-G-IT assay [73]. The available results seem to indicate that indeterminate results using QFN-G are age-dependent, being that IFN- γ is released in significantly lower quantities in response to phytohemagglutinin in young children [72,92]. By contrast, T-SPOT.TB, except in the first weeks of life [81], seems not to be age-dependent [12]. Nevertheless, Nicol *et al.* [93] reported a decrease in the number of children below the age of 1 year who scored positive T-SPOT.TB, whereas the TST results were unaffected by age. On the other hand, the QFN-G-IT test seems to offer a lower number of indeterminate results than the QFN-G test [75,76]. Indeed, recently, Bergamini *et al.*, in a retrospective study involving 496 children, reported that indeterminate results were associated with younger age (<4 years of age) for both QFN-G and QFN-G-IT, but not for T-SPOT.TB [94]. Lewinsohn *et al.*, using an in-house IFN- γ -based test based on ELISA detection, found that young household contact children (<2 years old) produced IFN- γ responses comparable to adults [85]. Liebeschuetz *et al.* reported that ELISPOT was less affected by HIV co-infection, malnutrition or age under 3 years old, than the TST [22]. Probably one of the main reasons that can explain the indeterminate results is the impossibility of isolating a high number of T cells in children if less of 4 ml of blood is collected.

One unresolved issue with IFN- γ -based tests is reversions and conversions observed in contact patients [95,96]. Hill *et al.* conducted a longitudinal study in the Gambia comparing ELISPOT results with those of the TST [96]. They included 740 contacts of 177 TB patients, all contacts being at least 15 years old. 3 and 18 months after the moment of recruitment, ELISPOT was repeated in contact patients. 3 months after recruitment

ELISPOT reversion was detected in 40% (54 out of 134) of contacts with a previous positive ELISPOT, and 18 months after recruitment reversion occurred in 36% (28 out of 78). On the contrary, the ELISPOT conversion rate was 27% (20 out of 75) at 18 months. Intriguingly, the rate of TST reversion was lower than in the ELISPOT, and the rate of TST conversion at 18 months was higher (50%). One plausible explanation is that reversion might reflect the clearance of bacilli from the organism [95], or might reflect the transition into latency [96]. In addition, the different rate of conversion between the ELISPOT and TST may also be due to a combination of factors: increased TST sensitivity, a different interval between initial exposure and test conversion, boosting the effect of TST results, and early ELISPOT reversion [96]. Therefore, more studies are needed to determine the biological variability of IFN- γ responses over time in the absence of TB exposure, and also to define the exact criteria for ELISPOT reversion and conversion. Further research is required for better understanding the meaning of these reversions and conversions [97]. All these facts should be considered when preparing IFN- γ -based test guidelines.

Bakir *et al.*, in a study conducted in Turkey involving more than 900 children and adolescents with recent household TB exposure, found that among the 381 contacts with positive ELISPOT, 11 developed active TB, and among the 550 with positive TST, 12 developed active TB [98]. They concluded that a positive ELISPOT result predicted the development of active-TB as well as the TST.

In children, IFN- γ -based tests have demonstrated to be useful in diagnosing LTBI, showing association between positive ELISPOT results with exposure to *M. tuberculosis* and not being affected by BCG vaccination. They could also be a useful tool for helping in the diagnosis of active TB. It is necessary to cautiously interpret a positive TST in screening studies, as the high ratio of discordant results with IFN- γ -based tests is probably due to NTM infection.

Diagnosing active TB

Some studies [12,13,23,30,34,99] and case reports [63,74,100–102] referring to the utility of IFN- γ -based tests in diagnosing active TB in immunocompromised patients have been reported. Nowadays, IFN- γ -based tests can be used as a complementary tool in the diagnosis of active TB, being helpful in areas where the prevalence of LTBI infection is very low. In addition, these tests can also be of use in diagnosing extrapulmonary TB, where approximately half of the cases have not been microbiologically diagnosed, and which often require complex and invasive diagnostic tools. Finally, tests could also be useful for pulmonary smear-negative patients. However, IFN- γ -based assays, as they are currently designed, cannot distinguish between active and latent TB infection.

Nevertheless, a novel approach has been proposed for diagnosing active TB. It is based on applying ELISPOT in samples collected directly from the site of infection. The recruitment of specific T cells during active TB and how antigen-specific cells clonally expand and migrate to the site of infection have been described [103].

Indeed, the utility of the ELISPOT in diagnosing active TB in immunocompromised patients using samples different from blood has already been reported. Richeldi *et al.* described how

an ELISPOT based on RD1 antigens on a pleural effusion sample accelerated the diagnosis of disseminated TB in a leukemic patient with persistent fever and a negative TST [104]. In this case, the ELISPOT on blood was positive 5 weeks before the TST became positive; and the ELISPOT on the pleural fluid sample was also positive 1.5 weeks before the mycobacteria isolated on the peripheral blood was identified as *M. tuberculosis*. Lange *et al.* also described the diagnosis of active TB in a patient receiving anti-TNF- α therapy by means of T-SPOT.TB on a pleural effusion sample [105]. The TST was negative (0 mm), the smear of sputum and bronchoalveolar lavage (BAL) fluid was also negative, and only the sputum culture and the pleural biopsy study (histopathology and nucleic acid-amplification technique) confirmed active TB. However, the sputum culture result was delayed between 7 and 35 days, and the nucleic acid-amplification technique on pleural biopsy was performed retrospectively.

By contrast, Baba *et al.* designed a study for validating the QFN-G assay on pleural fluid samples, in a group of HIV-infected patients with confirmed TB, probable TB and non-TB pleuritis [106]. The authors found that the QFN-G assay obtained 52% of indeterminate results, mainly caused by the high background observed in the negative control of both TB groups.

The detection of reactive T cells against the *M. tuberculosis*-specific antigens in BAL fluid by ELISPOT has been also described as compatible with the diagnosis of pulmonary active TB [107]. Regarding the experience in immunocompromised patients, its utility has been described in a patient receiving immunosuppressive therapies for a mixed connective tissue disease [108]. The patient had a negative TST, and smear and nucleic acid-amplification techniques were also negative in both the sputum and BAL fluid. The treatment was initiated, based on the ELISPOT result obtained from BAL fluid, 18 days before *M. tuberculosis* was isolated in the sputum culture.

Expert commentary

Since the development of IFN- γ -based assays as an *in vitro* alternative method to the TST for the immunodiagnosis of LTBI, promising results in adults and also in children have been published [12,70,72,76,79,80,109–111]. Although IFN- γ -based tests are designed for diagnosing LTBI, they have also been evaluated as an aid in the diagnosis of active TB [109,112–114]. A high specificity of the IFN- γ -based tests has been demonstrated, being that the IFN- γ -based tests were not affected by BCG vaccination or infection by the most common NTM. In addition, in the absence of a gold standard test to determine LTBI, IFN- γ test results have demonstrated to be closer than the TST to the degree of exposure to *M. tuberculosis* [80,100,115]. One of the key questions for its utilization is establishing the capacity of these tests in predicting the development of disease. In studies recently published [98,116,117], positive IFN- γ -based assays predicted development of active TB in adult and children with recent TB contact.

IFN- γ -based tests offer general advantages over the TST [118]: avoidance of cross-reaction with BCG-vaccinated individuals and with NTM infection, logistical convenience (result available in 24 h), avoidance of poorly reproducible measurements, no follow-up visit required and the result remains confidential. Furthermore,

an injection of PPD for the TST can boost subsequent TST responses, primarily in NTM-infected or BCG-vaccinated individuals. On the contrary, IFN- γ -based tests could be performed serially without inducing the boosting phenomenon.

According to cumulative evidence, the T-SPOT.TB assay showed a higher number of positive results, and a lower number of indeterminate results than the QFN tests. Given that the number of cells stimulated in the T-SPOT.TB instead of QFN test is standardized to 250,000 per culture, negative and indeterminate results associated with low lymphocyte counts could be reduced.

Nevertheless, although the T-SPOT.TB test seems to be more sensitive than the QFN-G-IT test, some operational aspects have to be noted for the performance of both IFN- γ -based tests (FIGURE 1). The T-SPOT.TB assay requires same-day processing of specimens, given the early and often inconvenient time limits for sample collection. This is especially important if a courier service is involved. Recently, the T-SPOT.TB manufacturer introduced a complementary reagent to allow the isolation of lymphocytes from whole blood up to 32 h following venopuncture. However, although the manufacturer claims that there is not a significant decrease in peripheral blood mononuclear cell yields or T-cell populations when comparing with whole-blood samples stored for less than 8 h, the procedure has not been extensively assessed and further experience is required.

Furthermore, T-SPOT.TB technical performance makes it difficult to run a large number of samples in a single day. In this sense, the QFN-G-IT test allows more flexible timing for specimen collection and transport owing to the fact that the antigens are already incorporated in the sample collection tube. The QFN-G-IT test also makes it possible to store the samples after stimulation with the antigens and to run batches. However, there are few studies evaluating whether stimulation of T cells with antigens together instead of separately could affect the sensitivity or specificity of the test. In a previous study evaluating LTBI in a rural area of South Africa, the QFN-G-IT test (201/358; 56%) obtained more positive results than the conventional QFN-G test (137/358; 38%) [119]. Furthermore, T-SPOT.TB requires more expertise in the readout of the results than QFN-G-IT, especially when the result is close to the cut-off value.

For immunosuppressed patients, additional advantages are required, such as a low ratio of indeterminate results, a high predictive value and a higher sensitivity than TST for avoiding false-negative results due to anergy.

Indeterminate results were mainly associated with immunosuppression, especially related to immunosuppressant therapy, and in patients with suppressed cellular immunity. Nevertheless, it is important to remark that in immunosuppressed patients with a negative TST result, an indeterminate IFN- γ test result should be considered as very useful clinical information.

The fact that IFN- γ -based tests measure effector T-cell responses rather than memory T cells seems to indicate that IFN- γ -based tests are a good indicator of recent LTBI, allowing the identification of patients with remote LTBI. Although this fact could be useful for selecting candidates to receive prophylaxis in an immunocompetent population, at the moment it is recommended to treat all immunocompromised patients with LTBI (both recent and remote infections).

Therefore, although among immunosuppressed patients the percentage of negative TST results is higher, and IFN- γ -based tests obtain more positive results with a higher specificity, the presence of discordant results between TST and IFN- γ -based tests, and the high risk of progression to active TB in this population, it seems prudent to recommend the utilization of IFN- γ -based tests after a negative TST result in order to increase the sensitivity of detecting LTBI cases in severely immunosuppressed patients. According to the data available, there is no risk of TST boosting in the IFN- γ -based test response if these tests are performed after a negative TST [120–124]. The possibility of using IFN- γ -based tests alone to avoid a large number of false-positive TST results should be considered for BCG-vaccinated nonsevere immunocompromised patients.

Although more studies comparing IFN- γ -based tests with the TST are needed to sufficiently expand knowledge, IFN- γ -based tests seem to be, in combination with the TST, a useful method for diagnosing TB infection in immunocompromised patients and children, according to their exact role in the different immunocompromising situations, the degrees of immunosuppression and the specific risks of LTBI for each patient.

Five-year view

In the next 5 years, the evolution of the IFN- γ -based tests in diagnosing TB infection will require advancements in two directions. First, it will be necessary to perform studies with a large number of patients to establish the prognostic value of positive test results in the different populations of immunosuppressed patients, especially in those that undergo an anti-TNF- α therapy.

Guidelines for IFN- γ -based test utilization in each group of immunocompromised patients and in each individual situation will be established. Given that *in vitro* assays depend on the secretion of IFN- γ , which is largely produced by CD4 T cells, more studies will be developed in order to define the CD4 threshold at which the performance of these assays declines, in particular when CD4 T-cell counts are under 50 cells/ μ l. These studies should explore the necessity and accuracy of new cut offs for diagnosing LTBI in immunosuppressed patients, and for distinguishing between active and latent TB infection.

Furthermore, specific studies will be performed in order to understand the discordant results between IFN- γ -based tests and the TST, especially in the pediatric population where the effect of NTM infection may play a very important role. The knowledge of the meaning of conversion and reversion results will also increase, and the clinical explanation of discordant and indeterminate results between IFN- γ -based tests will be expanded.

Second, technical modifications will be performed on IFN- γ -based tests [125]. The refinement of IFN- γ -based tests will include the exploration of alternative readouts to measure IFN- γ release [125], the utilization of new *M. tuberculosis*-specific antigens [6,126], and the simultaneous measurement of chemokines [127] and interleukins [128]. The next generation of IFN- γ -based tests will significantly enhance diagnostic sensitivity without diminishing specificity, and reduce the number of indeterminate results. In addition, the commercial IFN- γ -based test manufacturers should increase efforts to simplify technology and enhance its applicability in resource-limited settings.

Similarly, the detection of *M. tuberculosis*-specific T cells in nonblood samples using ELISPOT is a promising tool for the diagnosis of active TB in immunocompromised patients with negative smears. The definition of methodological procedures and the establishment of an accurate cut off will also be performed.

Financial & competing interests disclosure

The authors are members of the Tuberculosis Network European Trials group (TB-NET), Sociedad Española de Neumología y Cirugía Torácica, Societat Catalana de Pneumologia (SOCAP), Fundació Catalana de Pneumologia (FUCAP) and Instituto de Salud Carlos III (RETIC RD06/0018) have supported projects related to diagnosing active and latent TB infection by means of in vitro assays. Irene Latorre is a FPU pre-doctoral student and is the recipient of a grant from the 'Ministerio de Educación y Ciencia'. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

No writing assistance was utilized in the production of this manuscript.

Key issues

- IFN- γ -based tests produce more positive results than the tuberculin skin test (TST) for diagnosing latent TB infection (LTBI) in immunosuppressed patients, but less than in immunocompetent patients.
- Concordance between the TST and IFN- γ -based tests is poor, but IFN- γ -based test results are more likely associated with risk factors for LTBI than the TST.
- T-SPOT.TB shows a higher number of positive results than the QuantiFERON[®]-TB (QFN) test.
- Available data suggest that the T-SPOT.TB assay is less influenced by immunosuppression status than the QFN test.
- The T-SPOT.TB assay has a lower number of indeterminate results than the QFN test.
- Differences in the performance of both commercial tests should be considered when selecting the adequate IFN- γ -based test for each population and setting.
- In children, IFN- γ -based tests show a higher number of positive results than the TST, and have an especially higher specificity than the TST.
- The role of nontuberculous mycobacteria infection in children requires further studies for determining their effect in discordant results between the TST and IFN- γ -based tests.
- IFN- γ -based tests seem to be, in combination with the TST, a useful method for diagnosing TB infection in severely immunosuppressed patients.

References

Papers of special note have been highlighted as:

• of interest

•• of considerable interest

- 1 Global tuberculosis control: surveillance, planning, financing. WHO report. WHO, Geneva, Switzerland (WHO/HTM/TB/2005.349) (2005).
- 2 Jasmer RM, Nahid P, Hopewell PC. Clinical practice. Latent tuberculosis infection. *N. Engl. J. Med.* 347(23), 1860–1866 (2002).
- 3 Huebner RE, Schein MF, Bass JB Jr. The tuberculin skin test. *Clin. Infect. Dis.* 17(6), 968–975 (1993).
- 4 Horsburgh CR Jr. Priorities for the treatment of latent tuberculosis infection in the United States. *N. Engl. J. Med.* 350(20), 2060–2067 (2004).
- 5 Andersen P, Munk ME, Pollock JM, Doherty TM. Specific immune-based diagnosis of tuberculosis. *Lancet* 356(9235), 1099–1104 (2000).
- **Interesting description of the *Mycobacterium tuberculosis* specific antigens encoded in the RD1 region and its utility in immunodiagnosis.**
- 6 Brock I, Weldingh K, Leyten EM *et al.* Specific T-cell epitopes for immunoassay-based diagnosis of *Mycobacterium tuberculosis* infection. *J. Clin. Microbiol.* 42(6), 2379–2387 (2004).
- 7 Lalvani A, Millington KA. T cell-based diagnosis of childhood tuberculosis infection. *Curr. Opin. Infect. Dis.* 20(3), 264–271 (2007).
- 8 National Collaborating Centre for Chronic Conditions. *Tuberculosis: Clinical Diagnosis and Management of Tuberculosis, and Measures for its Prevention and Control*. Royal College of Physicians. Royal Imperial College, London, UK (2006).
- 9 Ruiz-Manzano J, Blanquer R, Calpe JL *et al.* SEPAR Guidelines. Diagnostic and treatment of tuberculosis. *Arch. Bronconeumol.* 44, 551–566 (2008).
- 10 Mazurek GH, Jereb J, Lobue P *et al.* Guidelines for using the QuantiFERON-TB Gold test for detecting *Mycobacterium tuberculosis* infection, United States. *MMWR Recomm. Rep.* 54(RR-15), 49–55 (2005).
- 11 Menzies D, Pai M, Comstock G. Meta-analysis: new tests for the diagnosis of latent tuberculosis infection: areas of uncertainty and recommendations for research. *Ann. Intern. Med.* 146(5), 340–354 (2007).
- **Comprehensive systematic review of the clinical utility of IFN- γ -based tests for diagnosing latent TB infection (LTBI), highlighting the issues that need further research.**
- 12 Ferrara G, Losi M, D'Amico R *et al.* Use in routine clinical practice of two commercial blood tests for diagnosis of infection with *Mycobacterium tuberculosis*: a prospective study. *Lancet* 367(9519), 1328–1334 (2006).
- **Utility of the *in vitro* tests in routine clinical practice, describing the impact of immunosuppression in indeterminate results.**
- 13 Kobashi Y, Mouri K, Obase Y *et al.* Clinical evaluation of QuantiFERON TB-2G test for immunocompromised patients. *Eur. Respir. J.* 30(5), 945–950 (2007).
- 14 Richeldi L, Losi M, D'Amico R *et al.* Performance of tests for latent tuberculosis in different groups of immunocompromised patients. *Chest* DOI:10.1178/chest.08–2575 (2009) (Epub ahead of print).
- 15 Duncan LE, Elliott AM, Hayes RJ *et al.* Tuberculin sensitivity and HIV-1 status of patients attending a sexually transmitted diseases clinic in Lusaka, Zambia: a cross-sectional study. *Trans. R. Soc. Trop. Med. Hyg.* 89(1), 37–40 (1995).
- 16 Markowitz N, Hansen NI, Wilcosky TC *et al.* Tuberculin and anergy testing in HIV-seropositive and HIV-seronegative persons. Pulmonary complications of HIV Infection Study Group. *Ann. Intern. Med.* 119(3), 185–193 (1993).
- 17 Rangaka MX, Wilkinson KA, Seldon R *et al.* Effect of HIV-1 infection on T-cell-based and skin test detection of tuberculosis infection. *Am. J. Respir. Crit. Care Med.* 175(5), 514–520 (2007).
- 18 Mandalakas AM, Hesselting AC, Chegou NN *et al.* High level of discordant IGRA results in HIV-infected adults and children. *Int. J. Tuberc. Lung Dis.* 12(4), 417–423 (2008).
- 19 Karam F, Mbow F, Fletcher H *et al.* Sensitivity of IFN- γ release assay to detect latent tuberculosis infection is retained in HIV-infected patients but dependent on HIV/AIDS progression. *PLoS ONE* 3(1), e1441 (2008).
- 20 Luetkemeyer AF, Charlebois ED, Flores LL *et al.* Comparison of an interferon- γ release assay with tuberculin skin testing in HIV-infected individuals. *Am. J. Respir. Crit. Care Med.* 175(7), 737–742 (2007).
- 21 Talati NJ, Seybold U, Humphrey B *et al.* Poor concordance between interferon- γ release assays and tuberculin skin tests in diagnosis of latent tuberculosis infection among HIV-infected individuals. *BMC Infect. Dis.* 9, 15 (2009).
- 22 Liebeschuetz S, Bamber S, Ewer K *et al.* Diagnosis of tuberculosis in South African children with a T-cell-based assay: a prospective cohort study. *Lancet* 364(9452), 2196–2203 (2004).
- 23 Vincenti D, Carrara S, Butera O *et al.* Response to region of difference 1 (RD1) epitopes in human immunodeficiency virus (HIV)-infected individuals enrolled with suspected active tuberculosis: a pilot study. *Clin. Exp. Immunol.* 150(1), 91–98 (2007).
- 24 Brock I, Ruhwald M, Lundgren B *et al.* Latent tuberculosis in HIV positive, diagnosed by the *M. tuberculosis* specific interferon- γ test. *Respir. Res.* 7, 56 (2006).
- 25 Balcells ME, Perez CM, Chanqueo L *et al.* A comparative study of two different methods for the detection of latent tuberculosis in HIV-positive individuals in Chile. *Int. J. Infect. Dis.* 12(6), 645–652 (2008).
- 26 Jones S, de Gijzel D, Wallach FR *et al.* Utility of QuantiFERON-TB Gold in-tube testing for latent TB infection in HIV-infected individuals. *Int. J. Tuberc. Lung Dis.* 11(11), 1190–1195 (2007).
- 27 Stephan C, Wolf T, Goetsch U *et al.* Comparing QuantiFERON-tuberculosis gold, T-SPOT tuberculosis and tuberculin skin test in HIV-infected individuals from a low prevalence tuberculosis country. *AIDS* 22(18), 2471–2479 (2008).
- 28 Chapman AL, Munkanta M, Wilkinson KA *et al.* Rapid detection of active and latent tuberculosis infection in HIV-positive individuals by enumeration of *Mycobacterium tuberculosis*-specific T cells. *AIDS* 16(17), 2285–2293 (2002).
- 29 Dheda K, Lalvani A, Miller RF *et al.* Performance of a T-cell-based diagnostic test for tuberculosis infection in HIV-infected individuals is independent of CD4 cell count. *AIDS* 19(17), 2038–2041 (2005).
- 30 Clark SA, Martin SL, Pozniak A *et al.* Tuberculosis antigen-specific immune responses can be detected using enzyme-linked immunosorbent assay technology in human immunodeficiency virus (HIV)-1 patients with advanced disease. *Clin. Exp. Immunol.* 150(2), 238–244 (2007).
- 31 Lawn SD, Bangani N, Vogt M *et al.* Utility of interferon- γ ELISPOT assay response in highly tuberculosis-exposed patients with advanced HIV infection in South Africa. *BMC Infect. Dis.* 7, 99 (2007).

- 32 Hammond AS, McConkey SJ, Hill PC *et al.* Mycobacterial T cell responses in HIV-infected patients with advanced immunosuppression. *J. Infect. Dis.* 197(2), 295–299 (2008).
- 33 Rivas I, Latorre I, Sanvisens A *et al.* Prospective evaluation of latent tuberculosis with interferon- γ release assays in drug and alcohol abusers. *Epidemiol. Infect.* 1–6 (2009).
- 34 Raby E, Moyo M, Devendra A *et al.* The effects of HIV on the sensitivity of a whole blood IFN- γ release assay in Zambian adults with active tuberculosis. *PLoS ONE* 3(6), e2489 (2008).
- 35 Aabye MG, Ravn P, PrayGod G *et al.* The impact of HIV infection and CD4 cell count on the performance of an interferon γ release assay in patients with pulmonary tuberculosis. *PLoS ONE* 4(1), e4220 (2009).
- 36 Rangaka MX, Diwakar L, Seldon R *et al.* Clinical, immunological, and epidemiological importance of antituberculosis T cell responses in HIV-infected Africans. *Clin. Infect. Dis.* 44(12), 1639–1646 (2007).
- 37 Aichelburg MC, Rieger A, Breitenacker F *et al.* Detection and prediction of active tuberculosis disease by a whole-blood interferon- γ release assay in HIV-1-infected individuals. *Clin. Infect. Dis.* 48(7), 954–962 (2009).
- 38 Panes J, Gomollon F, Taxonera C *et al.* Crohn's disease: a review of current treatment with a focus on biologics. *Drugs* 67(17), 2511–2537 (2007).
- This is an excellent and comprehensive review of Crohn's disease, with emphasis on biological treatments and its risk for developing TB.
- 39 Gomez-Reino JJ, Carmona L, Valverde VR, Mola EM, Montero MD. Treatment of rheumatoid arthritis with tumor necrosis factor inhibitors may predispose to significant increase in tuberculosis risk: a multicenter active-surveillance report. *Arthritis Rheum.* 48(8), 2122–2127 (2003).
- 40 Gardam MA, Keystone EC, Menzies R *et al.* Anti-tumour necrosis factor agents and tuberculosis risk: mechanisms of action and clinical management. *Lancet Infect. Dis.* 3(3), 148–155 (2003).
- 41 Keane J, Gershon S, Wise RP *et al.* Tuberculosis associated with infliximab, a tumor necrosis factor α -neutralizing agent. *N. Engl. J. Med.* 345(15), 1098–1104 (2001).
- 42 Carmona L, Gomez-Reino JJ, Rodriguez-Valverde V *et al.* Effectiveness of recommendations to prevent reactivation of latent tuberculosis infection in patients treated with tumor necrosis factor antagonists. *Arthritis Rheum.* 52(6), 1766–1772 (2005).
- 43 Lalvani A, Millington KA. Screening for tuberculosis infection prior to initiation of anti-TNF therapy. *Autoimmun. Rev.* 8(2), 147–152 (2008).
- 44 Domínguez J, Latorre I. Role of the T-cell interferon- γ release assays in preventing reactivation of latent tuberculosis infection in immunosuppressed patients in treatment with anti-TNF agents. *J. Crohn's & Colitis* 2, 250–254 (2008).
- 45 Bocchino M, Matarese A, Bellofiore B *et al.* Performance of two commercial blood IFN- γ release assays for the detection of *Mycobacterium tuberculosis* infection in patient candidates for anti-TNF- α treatment. *Eur. J. Clin. Microbiol. Infect. Dis.* 27(10), 907–913 (2008).
- 46 Cobanoglu N, Ozcelik U, Kalyoncu U *et al.* Interferon- γ assays for the diagnosis of tuberculosis infection before using tumour necrosis factor- α blockers. *Int. J. Tuberc. Lung Dis.* 11(11), 1177–1182 (2007).
- 47 Ponce de Leon D, Acevedo-Vasquez E, Alvizuri S *et al.* Comparison of an interferon- γ assay with tuberculin skin testing for detection of tuberculosis (TB) infection in patients with rheumatoid arthritis in a TB-endemic population. *J. Rheumatol.* 35(5), 776–781 (2008).
- 48 Vassilopoulos D, Stamoulis N, Hadziyannis E, Archimandritis AJ. Usefulness of enzyme-linked immunospot assay (Elispot) compared to tuberculin skin testing for latent tuberculosis screening in rheumatic patients scheduled for anti-tumor necrosis factor treatment. *J. Rheumatol.* 35(7), 1271–1276 (2008).
- 49 Matulis G, Juni P, Villiger PM, Gadola SD. Detection of latent tuberculosis in immunosuppressed patients with autoimmune diseases: performance of a *Mycobacterium tuberculosis* antigen-specific interferon γ assay. *Ann. Rheum. Dis.* 67(1), 84–90 (2008).
- 50 Behar SM, Shin DS, Maier A *et al.* Use of the T-SPOT.TB assay to detect latent tuberculosis infection among rheumatic disease patients on immunosuppressive therapy. *J. Rheumatol.* 36(3), 546–551 (2009).
- 51 Schoepfer AM, Flogerzi B, Fallegger S *et al.* Comparison of interferon- γ release assay versus tuberculin skin test for tuberculosis screening in inflammatory bowel disease. *Am. J. Gastroenterol.* 103(11), 2799–2806 (2008).
- 52 Sellam J, Hamdi H, Roy C *et al.* Comparison of *in vitro*-specific blood tests with tuberculin skin test for diagnosis of latent tuberculosis before anti-TNF therapy. *Ann. Rheum. Dis.* 66(12), 1610–1615 (2007).
- 53 Murakami S, Takeno M, Kirino Y *et al.* Screening of tuberculosis by interferon- γ assay before biologic therapy for rheumatoid arthritis. *Tuberculosis (Edinb.)* 89(2), 136–141 (2009).
- 54 Martin J, Walsh C, Gibbs A *et al.* Comparison of interferon- γ -release assays and conventional screening tests before tumour necrosis factor- α blockade in patients with inflammatory arthritis. *Ann. Rheum. Dis.* DOI:10.1136/ard.2008.101857 (2009) (Epub ahead of print).
- 55 Takahashi H, Shigehara K, Yamamoto M *et al.* Interferon γ assay for detecting latent tuberculosis infection in rheumatoid arthritis patients during infliximab administration. *Rheumatol. Int.* 27(12), 1143–1148 (2007).
- 56 Greenberg JD, Reddy SM, Schloss SG *et al.* Comparison of an *in vitro* tuberculosis interferon- γ assay with delayed-type hypersensitivity testing for detection of latent *Mycobacterium tuberculosis*: a pilot study in rheumatoid arthritis. *J. Rheumatol.* 35(5), 770–775 (2008).
- 57 Bartalesi F, Vicidomini S, Goletti D *et al.* QuantiFERON-TB Gold and the TST are both useful for latent tuberculosis infection screening in autoimmune diseases. *Eur. Respir. J.* 33(3), 586–593 (2009).
- 58 Hamdi H, Mariette X, Godot V *et al.* Inhibition of anti-tuberculosis T-lymphocyte function with tumour necrosis factor antagonists. *Arthritis Res. Ther.* 8(4), R114 (2006).
- 59 Chen DY, Shen GH, Hsieh TY, Hsieh CW, Lan JL. Effectiveness of the combination of a whole-blood interferon- γ assay and the tuberculin skin test in detecting latent tuberculosis infection in rheumatoid arthritis patients receiving adalimumab therapy. *Arthritis Rheum.* 59(6), 800–806 (2008).
- 60 Pratt A, Nicholl K, Kay L. Use of the QuantiFERON TB Gold test as part of a screening programme in patients with RA under consideration for treatment with anti-TNF- α agents: the Newcastle (UK) experience. *Rheumatology (Oxford)* 46(6), 1035–1036 (2007).
- 61 Munoz P, Rodriguez C, Bouza E. *Mycobacterium tuberculosis* infection in recipients of solid organ transplants. *Clin. Infect. Dis.* 40(4), 581–587 (2005).

- 62 Manuel O, Humar A, Preiksaitis J *et al.* Comparison of QuantiFERON-TB Gold with tuberculin skin test for detecting latent tuberculosis infection prior to liver transplantation. *Am. J. Transplant.* 7(12), 2797–2801 (2007).
- 63 Codeluppi M, Coechi S, Guaraldi G *et al.* Posttransplant *Mycobacterium tuberculosis* disease following liver transplantation and the need for cautious evaluation of QuantiFERON TB GOLD results in the transplant setting: a case report. *Transplant. Proc.* 38(4), 1083–1085 (2006).
- 64 Barsegian V, Mathias KD, Wrighton-Smith P, Grosse-Wilde H, Lindemann M. Prevalence of latent tuberculosis infection in German radiologists. *J. Hosp. Infect.* 69, 60–76 (2008).
- 65 Passalent L, Khan K, Richardson R *et al.* Detecting latent tuberculosis infection in hemodialysis patients: a head-to-head comparison of the T-SPOT.TB test, tuberculin skin test, and an expert physician panel. *Clin. J. Am. Soc. Nephrol.* 2(1), 68–73 (2007).
- 66 Winthrop KL, Nyendak M, Calvet H *et al.* Interferon- γ release assays for diagnosing *Mycobacterium tuberculosis* infection in renal dialysis patients. *Clin. J. Am. Soc. Nephrol.* 3(5), 1357–1363 (2008).
- 67 Triverio PA, Bridevaux PO, Roux-Lombard P *et al.* Interferon- γ release assays versus tuberculin skin testing for detection of latent tuberculosis in chronic haemodialysis patients. *Nephrol. Dial. Transplant.* DOI:10.1093/ndt/gfn748 (2009) (Epub ahead of print).
- 68 Lee SS, Chou KJ, Su IJ *et al.* High prevalence of latent tuberculosis infection in patients in end-stage renal disease on hemodialysis: comparison of QuantiFERON-TB GOLD, ELISPOT, and tuberculin skin test. *Infection* DOI:10.1007/s15010-008-8082-3 (2008) (Epub ahead of print).
- 69 Hursitoglu M, Cikrikcioglu MA, Tukek T *et al.* Acute effect of low-flux hemodialysis process on the results of the interferon- γ -based QuantiFERON-TB Gold In-Tube test in end-stage renal disease patients. *Transpl. Infect. Dis.* 11(1), 28–32 (2009).
- 70 Piana F, Codecasa LR, Cavallerio P *et al.* Use of a T-cell-based test for detection of tuberculosis infection among immunocompromised patients. *Eur. Respir. J.* 28(1), 31–34 (2006).
- 71 Nicol MP, Pienaar D, Wood K *et al.* Enzyme-linked immunospot assay responses to early secretory antigenic target 6, culture filtrate protein 10, and purified protein derivative among children with tuberculosis: implications for diagnosis and monitoring of therapy. *Clin. Infect. Dis.* 40(9), 1301–1308 (2005).
- 72 Connell TG, Curtis N, Ranganathan SC, Buttery JP. Performance of a whole blood interferon γ assay for detecting latent infection with *Mycobacterium tuberculosis* in children. *Thorax* 61(7), 616–620 (2006).
- 73 Connell TG, Ritz N, Paxton GA *et al.* A three-way comparison of tuberculin skin testing, QuantiFERON-TB gold and T-SPOT.TB in children. *PLoS ONE* 3(7), e2624 (2008).
- 74 Richeldi L, Ewer K, Losi M *et al.* T-cell-based diagnosis of neonatal multidrug-resistant latent tuberculosis infection. *Pediatrics* 119(1), e1–e5 (2007).
- 75 Dogra S, Narang P, Mendiratta DK *et al.* Comparison of a whole blood interferon- γ assay with tuberculin skin testing for the detection of tuberculosis infection in hospitalized children in rural India. *J. Infect.* 54, 267–276 (2007).
- 76 Dominguez J, Ruiz-Manzano J, De Souza-Galvao M *et al.* Comparison of two commercially available γ interferon blood tests for immunodiagnosis of tuberculosis. *Clin. Vaccine Immunol.* 15(1), 168–171 (2008).
- **A large size study comparing the results of T-SPOT.TB and QuantiFERON-TB Gold In Tube in both active TB and LTBI in the adult and child population.**
- 77 Kampmann B, Whittaker E, Williams A *et al.* Interferon- γ release assays do not identify more children with active TB than TST. *Eur. Respir. J.* DOI:10.1183/0903193600153408 (2009) (Epub ahead of print).
- 78 Davies MA, Connell T, Johannisen C *et al.* Detection of tuberculosis in HIV-infected children using an enzyme-linked immunospot assay. *AIDS* 23(8), 961–969 (2009).
- 79 Ewer K, Deeks J, Alvarez L *et al.* Comparison of T-cell-based assay with tuberculin skin test for diagnosis of *Mycobacterium tuberculosis* infection in a school tuberculosis outbreak. *Lancet* 361(9364), 1168–1173 (2003).
- **Very interesting study showing the association between positive ELISPOT results and exposure to *M. tuberculosis*.**
- 80 Lalvani A, Pathan AA, Durkan H *et al.* Enhanced contact tracing and spatial tracking of *Mycobacterium tuberculosis* infection by enumeration of antigen-specific T cells. *Lancet* 357(9273), 2017–2021 (2001).
- 81 Richeldi L, Ewer K, Losi M *et al.* T cell-based tracking of multidrug resistant tuberculosis infection after brief exposure. *Am. J. Respir. Crit. Care Med.* 170(3), 288–295 (2004).
- 82 Brock I, Welding K, Lillebaek T, Follmann F, Andersen P. Comparison of tuberculin skin test and new specific blood test in tuberculosis contacts. *Am. J. Respir. Crit. Care Med.* 170(1), 65–69 (2004).
- 83 Hesselting AC, Mandalakas AM, Kirchner HL *et al.* Highly discordant T-cell responses in individuals with recent household tuberculosis exposure. *Thorax* DOI:10.1136/thx.2007.085340 (2008) (Epub ahead of print).
- 84 Winje BA, Oftung F, Korsvold GE *et al.* School based screening for tuberculosis infection in Norway: comparison of positive tuberculin skin test with interferon- γ release assay. *BMC Infect. Dis.* 8, 140 (2008).
- **Provides new evidence of the role of nontuberculous mycobacteria in false-positive tuberculin skin tests in children screened for LTBI.**
- 85 Lewinsohn DA, Zalwango S, Stein CM *et al.* Whole blood interferon- γ responses to *Mycobacterium tuberculosis* antigens of persons with tuberculosis in Uganda. *PLoS ONE* 3(10), e3407 (2008).
- 86 Hill PC, Brookes RH, Adefifa MO *et al.* Comparison of enzyme-linked immunospot assay and tuberculin skin test in healthy children exposed to *Mycobacterium tuberculosis*. *Pediatrics* 117(5), 1542–1548 (2006).
- 87 Nakaoka H, Lawson L, Squire SB *et al.* Risk for tuberculosis among children. *Emerg. Infect. Dis.* 12(9), 1383–1388 (2006).
- 88 Chun JK, Kim CK, Kim HS *et al.* The role of a whole blood interferon- γ assay for the detection of latent tuberculosis infection in Bacille Calmette–Guerin vaccinated children. *Diagn. Microbiol. Infect. Dis.* 62(4), 389–394 (2008).
- 89 de Jong BC, Hill PC, Brookes RH *et al.* *Mycobacterium africanum* elicits an attenuated T cell response to early secreted antigenic target, 6kDa, in patients with tuberculosis and their household contacts. *J. Infect. Dis.* 193(9), 1279–1286 (2006).

- 90 Detjen AK, Keil T, Roll S *et al.* Interferon- γ release assays improve the diagnosis of tuberculosis and nontuberculous mycobacterial disease in children in a country with a low incidence of tuberculosis. *Clin. Infect. Dis.* 45(3), 322–328 (2007).
- 91 Hill PC, Jeffries DJ, Brookes RH *et al.* Using ELISPOT to expose false positive skin test conversion in tuberculosis contacts. *PLoS ONE* 2(1), e183 (2007).
- 92 Lighter J, Rigaud M, Eduardo R, Peng CH, Pollack H. Latent tuberculosis diagnosis in children by using the QuantiFERON-TB Gold In-Tube test. *Pediatrics* 123(1), 30–37 (2009).
- 93 Nicol MP, Davies MA, Wood K *et al.* Comparison of T-SPOT:TB assay and tuberculin skin test for the evaluation of young children at high risk for tuberculosis in a community setting. *Pediatrics* 123(1), 38–43 (2009).
- 94 Bergamini BM, Losi M, Vaianti F *et al.* Performance of commercial blood tests for the diagnosis of latent tuberculosis infection in children and adolescents. *Pediatrics* 123(3), e419–e424 (2009).
- 95 Ewer K, Millington KA, Deeks JJ *et al.* Dynamic antigen-specific T-cell responses after point-source exposure to *Mycobacterium tuberculosis*. *Am. J. Respir. Crit. Care Med.* 174(7), 831–839 (2006).
- 96 Hill PC, Brookes RH, Fox A *et al.* Longitudinal assessment of an ELISPOT test for *Mycobacterium tuberculosis* infection. *PLoS Med.* 4(6), e192 (2007).
- 97 Pai M, Dheda K, Cunningham J, Scano F, O'Brien R. T-cell assays for the diagnosis of latent tuberculosis infection: moving the research agenda forward. *Lancet Infect. Dis.* 7(6), 428–438 (2007).
- 98 Bakir M, Millington KA, Soysal A *et al.* Prognostic value of a T-cell-based, interferon- γ biomarker in children with tuberculosis contact. *Ann. Intern. Med.* 149(11), 777–787 (2008).
- 99 Kim SH, Song KH, Choi SJ *et al.* Diagnostic usefulness of a T-cell-based assay for extrapulmonary tuberculosis in immunocompromised patients. *Am. J. Med.* 122(2), 189–195 (2009).
- 100 Richeldi L, Ewer K, Losi M *et al.* Early diagnosis of subclinical multidrug-resistant tuberculosis. *Ann. Intern. Med.* 140(9), 709–713 (2004).
- 101 Spyridis N, Chakraborty R, Sharland M, Heath PT. Early diagnosis of tuberculosis using an INF- γ assay in a child with HIV-1 infection and a very low CD4 count. *Scand. J. Infect. Dis.* 39(10), 919–921 (2007).
- 102 Ravn P, Munk ME, Andersen AB *et al.* Reactivation of tuberculosis during immunosuppressive treatment in a patient with a positive QuantiFERON-RD1 test. *Scand. J. Infect. Dis.* 36(6–7), 499–501 (2004).
- 103 Wilkinson KA, Wilkinson RJ, Pathan A *et al.* *Ex vivo* characterization of early secretory antigenic target 6-specific T cells at sites of active disease in pleural tuberculosis. *Clin. Infect. Dis.* 40(1), 184–187 (2005).
- 104 Richeldi L, Luppi M, Losi M *et al.* Diagnosis of occult tuberculosis in hematological malignancy by enumeration of antigen-specific T cells. *Leukemia* 20(2), 379–381 (2006).
- 105 Lange C, Hellmich B, Ernst M, Ehlers S. Rapid immunodiagnosis of tuberculosis in a woman receiving anti-TNF therapy. *N. Clin. Pract. Rheumatol.* 3(9), 528–534 (2007).
- **This case report suggests that detection of T cells in bronchoalveolar lavage by T-SPOT:TB may be useful in diagnosing active TB.**
- 106 Baba K, Sornes S, Hoosen AA *et al.* Evaluation of immune responses in HIV infected patients with pleural tuberculosis by the QuantiFERON TB-Gold interferon- γ assay. *BMC Infect. Dis.* 8, 35 (2008).
- 107 Jafari C, Ernst M, Strassburg A *et al.* Local immunodiagnosis of pulmonary tuberculosis by enzyme-linked immunospot. *Eur. Respir. J.* 31(2), 261–265 (2008).
- 108 Strassburg A, Jafari C, Ernst M, Lotz W, Lange C. Rapid diagnosis of pulmonary TB by BAL enzyme-linked immunospot assay in an immunocompromised host. *Eur. Respir. J.* 31(5), 1132–1135 (2008).
- 109 Goletti D, Stefania C, Butera O *et al.* Accuracy of immunodiagnostic tests for active tuberculosis using single and combined results: a multicenter TBNET-Study. *PLoS ONE* 3(10), e3417 (2008).
- 110 Lalvani A, Nagvenkar P, Udwardia Z *et al.* Enumeration of T cells specific for RD1-encoded antigens suggests a high prevalence of latent *Mycobacterium tuberculosis* infection in healthy urban Indians. *J. Infect. Dis.* 183(3), 469–477 (2001).
- 111 Richeldi L, Losi M, Cerri S *et al.* Using ELISpot technology to improve the diagnosis of tuberculosis infection: from the bench to the T-SPOT:TB assay. *Expert Rev. Respir. Med.* 2(2), 253–260 (2007).
- 112 Ravn P, Munk ME, Andersen AB *et al.* Prospective evaluation of a whole-blood test using *Mycobacterium tuberculosis*-specific antigens ESAT-6 and CFP-10 for diagnosis of active tuberculosis. *Clin. Diagn. Lab. Immunol.* 12(4), 491–496 (2005).
- 113 Domínguez J, De Souza-Galvao M, Ruiz-Manzano J *et al.* T-cell responses to the *Mycobacterium tuberculosis*-specific antigens in active tuberculosis patients at the beginning, during, and after antituberculosis treatment. *Diagn. Microbiol. Infect. Dis.* 63, 43–51 (2009).
- 114 Bothamley G. Interferon- γ -release assays in the management of tuberculosis. *Expert Rev. Respir. Med.* 1(3), 365–375 (2007).
- 115 Soysal A, Millington KA, Bakir M *et al.* Effect of BCG vaccination on risk of *Mycobacterium tuberculosis* infection in children with household tuberculosis contact: a prospective community-based study. *Lancet* 366(9495), 1443–1451 (2005).
- 116 Hill PC, Jackson-Sillah D, Fox A *et al.* Incidence of tuberculosis and the predictive value of ELISPOT and Mantoux tests in Gambian case contacts. *PLoS ONE* 3(1), e1379 (2008).
- 117 Diel R, Loddenkemper R, Meywald-Walter K, Niemann S, Nienhaus A. Predictive value of a whole-blood IFN- γ assay for the development of active TB disease. *Am. J. Respir. Crit. Care Med.* 177(10), 1164–1170 (2008).
- 118 Domínguez J, Ruiz-Manzano J. The tuberculin skin test: time for a change? *Arch. Bronconeumol.* 42(2), 47–48 (2006).
- 119 Mahomed H, Hughes EJ, Hawkrige T *et al.* Comparison of Mantoux skin test with three generations of a whole blood IFN- γ assay for tuberculosis infection. *Int. J. Tuberc. Lung Dis.* 10(3), 310–316 (2006).
- 120 Richeldi L, Bergamini BM, Vaianti F. Prior tuberculin skin testing does not boost QuantiFERON-TB results in paediatric contacts. *Eur. Respir. J.* 32(2), 524–525 (2008).
- 121 Choi JC, Shin JW, Kim JY *et al.* The effect of previous tuberculin skin test on the follow-up examination on whole-blood interferon- γ assay in the screening for latent tuberculosis infection. *Chest* 133(6), 1415–1420 (2008).
- 122 Naser A, Naqvi S, Kampmann B. Evidence for boosting *Mycobacterium tuberculosis*-specific IFN- γ responses at 6 weeks following tuberculin skin testing. *Eur. Respir. J.* 29(6), 1282–1283 (2007).

- 123 Richeldi L, Ewer K, Losi M *et al.* Repeated tuberculin testing does not induce false positive ELISPOT results. *Thorax* 61(2), 180 (2006).
- 124 Leyten EM, Prins C, Bossink AW *et al.* Effect of tuberculin skin testing on a *Mycobacterium tuberculosis*-specific interferon- γ assay. *Eur. Respir. J.* 29(6), 1212–1216 (2007).
- 125 Lalvani A, Millington KA. T-cell interferon- γ release assays: can we do better? *Eur. Respir. J.* 32(6), 1428–1430 (2008).
- 126 Liu XQ, Dosanjh DP, Varia H *et al.* Evaluation of T-cell responses to novel RD1- and RD2-encoded *Mycobacterium tuberculosis* gene products for specific detection of human tuberculosis infection. *Infect. Immun.* 72(5), 2574–2581 (2004).
- 127 Ruhwald M, Bodmer T, Maier C *et al.* Evaluating the potential of IP-10 and MCP-2 as biomarkers for the diagnosis of tuberculosis. *Eur. Respir. J.* 32(6), 1607–1615 (2008).
- 128 Millington KA, Innes JA, Hackforth S *et al.* Dynamic relationship between IFN- γ and IL-2 profile of *Mycobacterium tuberculosis*-specific T cells and antigen load. *J. Immunol.* 178(8), 5217–5226 (2007).
- Irene Latorre
Servei de Microbiologia, Fundació Institut d'Investigació en Ciències de la Salut 'Germans Trias i Pujol', Carretera del Canyet s/n, 08916 Badalona, Barcelona, Spain
and,
Universitat Autònoma de Barcelona, Bellaterra, Spain
and,
CIBER Enfermedades Respiratorias, Instituto de Salud Carlos III, Badalona, Barcelona, Spain
Tel.: +34 934 978 894
Fax: +34 934 978 895
irelatorre@gmail.com
 - Neus Altet
Unidad de Prevención y Control de la Tuberculosis de Barcelona, Instituto Catalán de la Salud, Avda, Drassanes, 17–21, Barcelona, Spain
Tel.: +34 933 012 424
Fax: +34 933 186 186
naltet.bcn.ics@gencat.cat
 - Lourdes Mateo
Servei de Reumatologia, Hospital Universitari 'Germans Trias i Pujol', Carretera del Canyet s/n, 08916 Badalona, Barcelona, Spain
and,
Universitat Autònoma de Barcelona, Bellaterra, Spain
Tel.: +34 934 651 200
Fax: +34 934 978 895
lmateo.germantrias@gencat.cat
 - Malú De Souza-Galvão
Unidad de Prevención y Control de la Tuberculosis de Barcelona, Instituto Catalán de la Salud, Avda, Drassanes, 17–21, Barcelona, Spain
and,
Universitat Autònoma de Barcelona, Bellaterra, Spain
Tel.: +34 933 012 424
Fax: +34 933 186 186
maludesouzagalvao@gmail.com
 - Juan Ruiz-Manzano
Servei de Pneumologia, Hospital Universitari 'Germans Trias i Pujol', Carretera del Canyet s/n, 08916 Badalona, Barcelona, Spain
and,
Universitat Autònoma de Barcelona, Bellaterra, Spain
and,
CIBER Enfermedades Respiratorias, Instituto de Salud Carlos III, Badalona, Barcelona Spain
Tel.: +34 934 651 200
Fax: +34 934 978 895
jrui@separ.es
 - Vicente Ausina
Servei de Microbiologia, Hospital Universitari 'Germans Trias i Pujol', Carretera del Canyet s/n, 08916 Badalona, Barcelona, Spain
and,
Universitat Autònoma de Barcelona, Bellaterra, Spain
and,
CIBER Enfermedades Respiratorias, Instituto de Salud Carlos III, Badalona, Barcelona, Spain
Tel.: +34 934 978 894
Fax: +34 934 978 895
vausina.germantrias@gencat.cat

Affiliations

- Jose Domínguez
Servei de Microbiologia, Fundació Institut d'Investigació en Ciències de la Salut 'Germans Trias i Pujol', Carretera del Canyet s/n, 08916 Badalona, Barcelona, Spain
and,
Universitat Autònoma de Barcelona, Bellaterra, Spain
and,
CIBER Enfermedades Respiratorias, Instituto de Salud Carlos III, Badalona, Barcelona, Spain
Tel.: +34 934 978 894
Fax: +34 934 978 895
jadomb@gmail.com

